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A Chemical Synthesis of 3-Deoxy-D-manno-2-octulosonic Acid from D-mannose

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A chemical synthesis of Kdo was achieved via the condensation of a protected D-mannofuranose-5,6-cyclic sulfate with ethyl 1,3-dithiane-2-carboxylate, which is an efficient equivalent for installing an α -ketoester unit as a key step.

INTRODUCTION

2-Keto-3-deoxy-D-manno-octulosonic acid (Kdo, **1**) is a component in the outer membrane lipopolysaccharide (LPS) or lipo-oligosaccharide (LOS) produced by gram-negative bacteria.

Its chemical and enzymatic syntheses from D-arabinose and D-mannose have been reported by several groups.^[1–3] Among them, two-carbon elongation at the anomeric position of the starting D-mannose derivative is reliable.^[4] In the chemical synthesis from D-mannose, all groups put emphasis on the elaboration of the two-carbon unit, which is later converted to α -ketocarboxylic acid moiety.

Inspired by studies in which a nucleophilic substitution of cyclic sulfates with dithianyl compounds afforded corresponding carbon elongated products,^[4c,4e,5] we designed a different approach to Kdo, which was the treatment of cyclic sulfate **2** with glyoxylate equivalent **3** to give the two-carbon elongated product **4** at the C-6 position, which might be transformed to Kdo (Fig. 1). In this paper the synthesis of Kdo ammonium salt using the alkylation of the cyclic sulfate **2** at the C-6 position with the ethyl dithiane-2-carboxylate anion is described.

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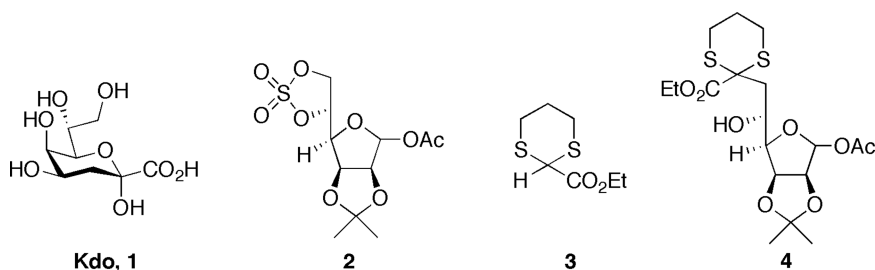
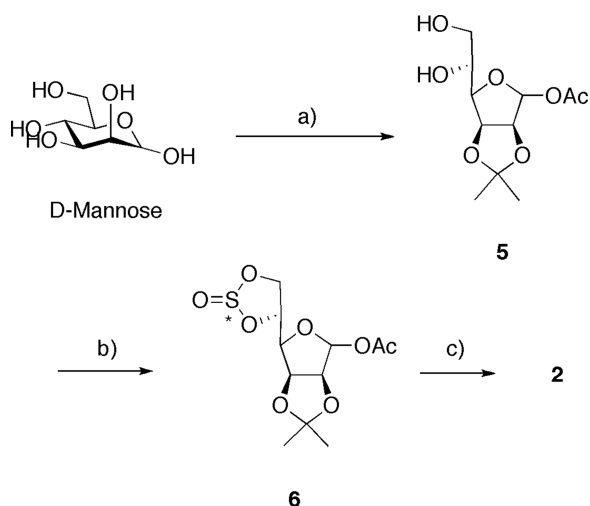


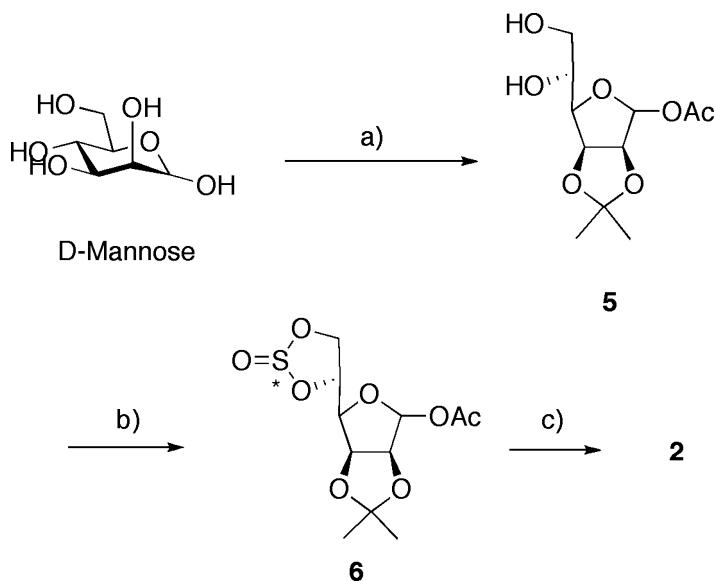
Figure 1:

RESULTS AND DISCUSSIONS

The key cyclic sulfate **2** for this study was prepared from diol **5**, which was easily synthesized from D-mannose in the following procedures (Scheme 1): acetalation of D-mannose,^[6] acetylation of the anomeric hydroxyl group,^[7] and acidic hydrolysis of 5,6-acetonide^[8] (>95% in three steps). Diol **5** was treated with thionyl chloride and triethylamine in dichloromethane to give cyclic sulfinites **6** in 99% yield as a 2:1 mixture of diastereomers. Although these diastereomers could be easily separated by silica gel column chromatography, they did not necessarily have to be separated due to the continuing oxidation to get the key cyclic sulfate **2**. Oxidation of the mixture of cyclic sulfites **6** with $\text{RuCl}_3 \cdot n\text{H}_2\text{O}$ and NaIO_4 ^[9] afforded cyclic sulfate **2** as a colorless solid in 92% yield. Compound **2** was stable under argon at -20°C for 6 months.



Scheme 1: Conditions: a) i) acetone, I_2 ,⁽⁶⁾ 99%, ii) Ac_2O , Py ,⁽⁷⁾ 99%, iii) 60% AcOH ,⁽⁸⁾ 99%; b) SOCl_2 , Et_3N , CH_2Cl_2 , 95%; c) $\text{RuCl}_3 \cdot n\text{H}_2\text{O}$, NaIO_4 , acetone, H_2O , 92%.



Scheme 2:

Ethyl 1,3-dithiane-2-carboxylate **3** was chosen as a suitable glyoxylate unit. The key alkylation of **2** with **3** was carried out as follows (Scheme 2). To a solution of the anion **3** (prepared with NaHMDS or NaH and a catalytic amount of *t*-butanol at 0°C) was added the cyclic sulfate **2**. The mixture was stirred at rt for 3 h, neutralized by the addition of a 1.0 equiv. of sulfuric acid and water, and separated by size exclusion chromatography (Sephadex LH-20, chloroform-methanol) and purified by silica gel column chromatography to give sulfate **7** and sulfate hemiacetal **7a**. Acid hydrolysis of **7** and **7a** gave a mixture of hydroxy ester **8**, lactone **9** (38%, 23% from **2**), and trace amounts of hemiacetals (**8a** and **9a**). Although these compounds could be separated by silica gel column chromatography, they converged into lactone **9** after acetylation of the anomeric hydroxyl group and intramolecular lactonization. Accordingly, compound **9** was obtained in 64% yield from cyclic sulfate **2** by sequentially carrying out acid hydrolysis, acetylation, and subsequent lactone formation without isolating **7**, **8**, and **9**.

The structures of **8** and **9** were determined by 2-D NMR spectroscopy (DQF-COSY, HMQC, and HMBC). The assignments of ring proton and carbon signals of both **8** and **9** were confirmed by comparing their NMR data to those of 1,5,6-tri-*O*-acetyl-2,3-*O*-isopropylidene-mannofuranose.^[10]

The structure of hydroxy ester **8** was determined by confirming that the carbon atom at C-9 and the quaternary carbon of dithioacetal are connected. In the HMBC spectrum, the presence of the cross-relay peak due to C-1 and the methylene protons of the ethyl group confirmed that the ¹³C satellite signal

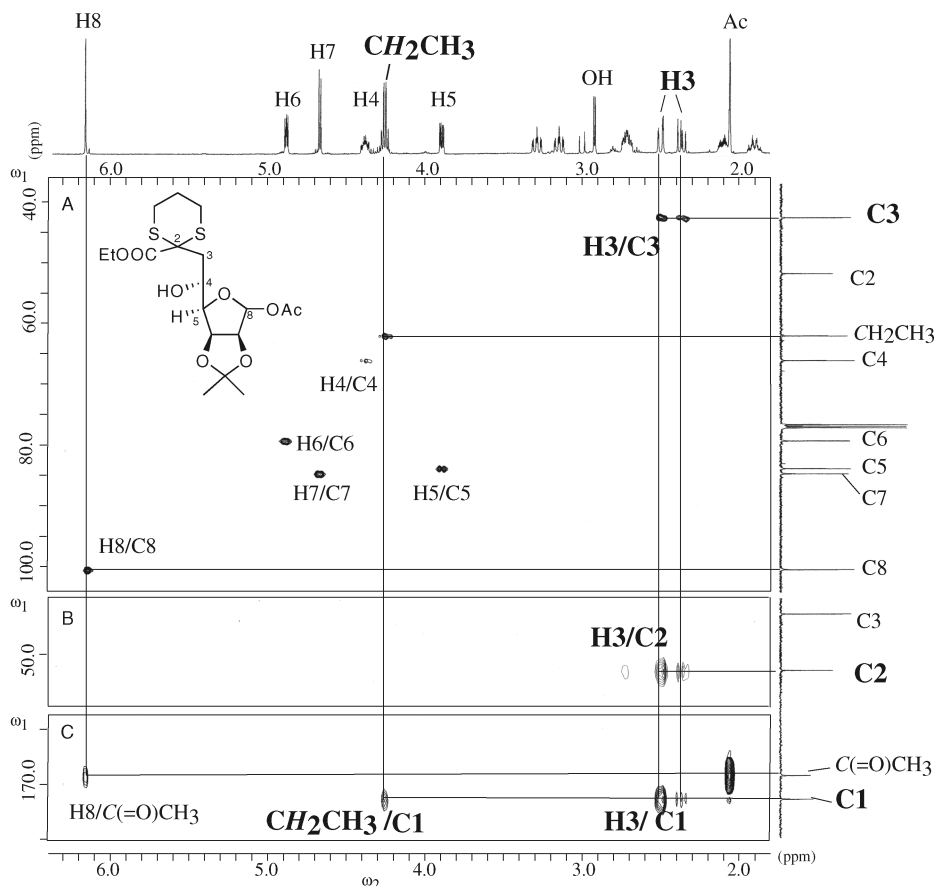


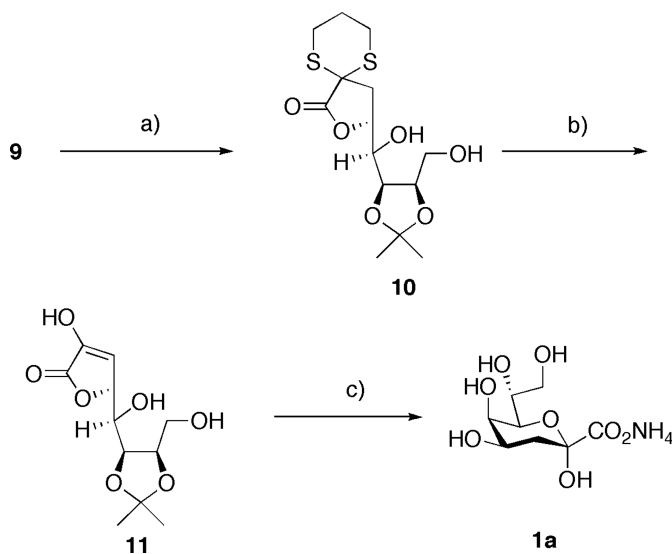
Figure 2: Partial HMQC (panel A) and HMBC (panels B and C) spectra of compound **8** recorded in CDCl_3 at 25°C . Only partial $^{13}\text{C}/^1\text{H}$ cross-peaks (A) and cross-relay peaks (B and C) are labeled.

at 169.2 ppm is C-1. In addition, the presence of two sets of cross-relay peaks ascribed to H-3/C-1 and H-3/the quaternary carbon of the dithiane group (C-2) was confirmed (Fig. 2). Thus, we determined that a dithiane **3** is connected to a cyclic sulfate **2** at the C-6 position.

In a similar manner, the absence of an ethyl signal due to COOEt in the ^1H NMR spectrum confirmed that the intramolecular cyclization took place to form a lactone structure. Therefore, we determined the structure of lactone **9**.

The deacetylation of lactone **9** with NaOMe and the reductive ring opening of the resulting hemiacetal with sodium borohydride (NaBH_4) at 0°C gave diol **10** in 86%. When the reduction was performed at rt, an undesired triol was given in 90% yield due to the reduction of the lactone function. The treatment

of **10** with N-bromosuccinimide (NBS) in aqueous acetone gave unsaturated lactone **11** in 78%. The isopropylidene group of **11** was cleaved with aqueous trifluoroacetic acid to give Kdo **1**, which was purified by ion-exchange column chromatography as its ammonium salt in 70% yield (Scheme 3).



Scheme 3: Conditions: a) NaOMe, MeOH, rt, then NaBH₄, 0°C, 86%; b) NBS, acetone, H₂O, 0°C, 78%; c) 90% TFA aq, rt, 15 min, 70%.

In summary, we successfully synthesized Kdo ammonium salt from easily available 1-*O*-acetyl-2,3-*O*-isopropylidene- α -D-mannofuranose in seven steps (overall 26%). This methodology for installing a glyoxylate dithioacetal unit onto the C-6 of D-mannose is convenient from the double points of view of the availability of the key α -ketoacid synthon (alkyl 1,3-dithiane-2-carboxylates are commercially available) and of the ready access to the desired ketoside carboxylic acid in four steps from the protected D-mannofuranose cyclic sulfate **2**. Furthermore, the synthetic intermediate **10** would be expected to convert into an acceptor that is suitable for the oligosaccharide synthesis, because it has two hydroxyl groups at the C-5 and C-8 positions, and each OH group will be able to connect with mono- or oligosaccharides to build inner core oligosaccharides of LOS/LPS. We are currently exploiting this intermediate **9** for the synthesis of core oligosaccharides of LOS as a glycosyl acceptor and will report the results in due course.

EXPERIMENTAL

General Methods

Optical rotations and melting points (uncorrected) were measured with a HORIBA SEPA-200 polarimeter and a YANAGIMOTO micro melting point apparatus, respectively. All NMR spectra were recorded at 25°C in CDCl₃ using a JEOL JNM-ECP 500-MHz NMR spectrometer equipped with a Silicon Graphics O₂ computer. Chemical shifts are reported in ppm relative to internal Me₄Si (δ_H 0.00) for ¹H NMR and CDCl₃ (δ_C 77.00) for ¹³C NMR. Two-dimensional NMR data (DQF-COSY, HMQC, and HMBC) were processed using a Delta program (JEOL USA Inc.) in a similar manner as described previously.^[11–13] Silica gel 60 (E. Merck) was used for flash column (0.040–0.063 mm) and open column (0.063–0.200 mm) chromatography. Silica gel 60 F₂₅₄ (E. Merck) was used for thin-layer chromatography (TLC), and compounds were detected under UV light (254 nm) or by spraying with 10% conc. H₂SO₄ in MeOH and then heating the plates at 120°C for 5 min.

1-*O*-acetyl-2,3-*O*-isopropylidene-5,6-sulfonylidene- α -D-mannofuranose (**2**)

To a solution of 1-*O*-acetyl-2,3-*O*-isopropylidenemannofuranose^[8] **5** (1.70 g, 6.48 mmol) and triethylamine (3.6 mL, 26 mmol) in dry dichloromethane (25 mL), a solution of thionylchloride (0.90 mL, 13.0 mmol) was added at 0°C. After being stirred for 2 h, the mixture was diluted with EtOAc and added to saturated aqueous NaHCO₃. The aqueous layer was extracted with three portions of EtOAc. The combined organic layers were washed with brine, dried over Na₂SO₄, filtered, and concentrated in vacuo. The residue was chromatographed on a column of silica gel (EtOAc:hexane = 3:2) to give a mixture of diastereomers as a colorless syrup **6** (1.99 g, 99%). A mixture of diastereomer was dissolved in a mixed solution of acetonitrile-dichloromethane-water (2:2:3, 49 mL), and then NaIO₄ (2.77 g, 13.0 mmol) and a catalytic amount of RuCl₃ · 7 H₂O (6.0 mg) was added at rt. The reaction mixture was vigorously stirred for 20 min and then diluted with ethyl acetate and water. The aqueous layer was extracted with three portions of ethyl acetate. The combined organic layers were washed with water and brine, dried over Na₂SO₄, and concentrated. The residue was chromatographed on a column of silica gel (hexane:ethyl acetate = 1:1) to afford a colorless solid, which was recrystallized from dichloromethane (1.93 g, 92%).

M.p. 90.3–91.3°C, $[\alpha]_D^{26} + 49.0$ (*c* 1.0, CHCl₃). ¹H NMR (CDCl₃) δ 6.20 (brs, 1H, H-1), 5.36 (ddd, 1H, $J_{4,5} = 6.0$, $J_{5,6a} = 6.5$, $J_{5,6b} = 6.5$ Hz, H-5), 4.90 (dd, 1H, $J_{2,3} = 5.5$, $J_{3,4} = 3.5$ Hz, H-3) 4.81 (dd, 1H, $J_{6a,6b} = 9.0$ Hz, H-6a), 4.77 (dd, 1H, H-6b), 4.75 (d, 1H, H-2), 4.49 (dd, 1H, H-4), 2.10, (s, 3H, COCH₃), 1.48 and

1.33 (s each, 3H, CH₃). ¹³C NMR (CDCl₃) δ 169.0 (COCH₃) 100.3 (C-1), 84.8 (C-2), 79.5 (C-4), 78.7 (C-3), 77.7 (C-5), 69.8 (C-6), 25.6 (CH₃), 24.1 (CH₃), 20.9 (COCH₃). Anal. calcd for C₁₁H₁₆O₉S; C, 40.74; H, 4.97; S, 9.89. Found C, 40.55; H, 4.85; S, 9.70.

(1S,2R,4R,5S,1'R)-4-{2'-[2-(ethoxycarbonyl)(1,3-dithian-2-yl)]-1'-hydroxyethyl}-7,7-dimethyl-3,6,8-trioxabicyclo[3.3.0]oct-2-yl acetate (8) and (1S,2R,4R,5S,3'R)-7,7-dimethyl-3,6,8-trioxa-4-(3'-oxa-4'-oxo-6',10'-dithiaspiro[4.5]dec-2'-yl)bicyclo[3.3.0]oct-2-yl acetate (9)

Method A: To a solution of NaHMDS (0.6 M, 6.0 mL, 3.6 mmol) in dry DMF (10 mL), a solution of 1,3-dithiane-2-carboxylate (593 mg, 3.1 mmol) in DMF (5.0 mL) was added at 0°C. After stirring the mixture for 20 min, a solution of cyclic sulfonate **2** (500 mg, 1.54 mmol) in DMF (5.0 mL) was added, and the mixture was warmed up to rt. After being stirred for 3 h, 1 equiv. of conc. H₂SO₄ and water (1 equiv.) was added at 0°C, and the mixture was concentrated in vacuo. The residue was chromatographed on a column of Sephadex LH-20 (CHCl₃:MeOH = 3:1) to give corresponding sulfate esters **7** and **7a** as a colorless powder. The powder was dissolved in 1,4-dioxane (10.0 mL) and treated with a 100 μ L of conc. H₂SO₄ and 100 μ L of water at 0°C for 1 h. The mixture was neutralized with sat. NaHCO₃ aq. and diluted with ethyl acetate and filtrated through celite, and the filtrate was concentrated in vacuo. The residue was chromatographed on a column of silica gel (hexane:ethyl acetate = 1:1) to afford compounds **8** (255 mg, 38%) and **9** (140 mg, 23%).

Method B: To a suspension of NaH (258 mg, 6.17 mmol), t-BuOH (10 μ L), and DMF (5 mL), a solution of compound **3** (1.19 g, 6.17 mmol) in DMF (7 mL) was added at 0°C. After stirring for 1 h, a solution of compound **2** (1.00 g, 3.08 mmol) in DMF (12 mL) was added to the mixture and it was stirred for 1.5 h. Sulfuric acid (320 μ L) and water (110 μ L) was added and then the reaction mixture was concentrated to remove DMF. The residue was dissolved in 1,4-dioxane (40 mL) and treated with sulfuric acid (650 μ L) and water (220 μ L) for 1 h at 0°C. The mixture was neutralized with sat. NaHCO₃ aq., diluted with dichloromethane, and separated. The aqueous solution was extracted with dichloromethane, and the combined extracts were washed with brine, dried (Na₂SO₄), filtered, and concentrated to a syrup. The residue was dissolved in pyridine (2.3 mL) and acetic anhydride (2.3 mL) and stirred for 12 h. The mixture was coevaporated with toluene several times to a residue. Purification by flash column chromatography (dichloromethane:hexane:ethyl acetate = 16:3:1) of the residue gave **9** as a colorless syrup (769 mg, 64%).
Compound 8: [α]_D²⁵+34.0 (c 1.0, CHCl₃). ¹H NMR (CDCl₃) δ 6.16 (brs, 1H, H-1), 4.89 (dd, 1H, *J*_{5,6} = 3.5, *J*_{6,7} = 6.0 Hz, H-6), 4.67, (brd, 1H, H-7), 4.34

(ddd, 1H, $J_{3a,4} = 9.5$, $J_{3b,4} = 2.0$, $J_{4,5} = 8.5$ Hz, H-4), 4.26 (q, 2H, $J = 7.0$ Hz, CH_2CH_3), 3.90 (dd, 1H, H-5), 3.32–3.27 and 3.18–3.13 (m, 1H each, SCH_2), 2.49 (ddd, 1H, $J_{3a,3b} = 14.5$ Hz, H-3a), 2.39 (ddd, 1H, H-3b), 2.15–2.08 (m, 2H, CH_2), 2.07 (s, 3H, Ac), 1.95–1.87 (m, 2H, CH_2), 1.48 and 1.33 (s, 3H each, CH_3), 1.34 (t, 3H, CH_2CH_3). ^{13}C NMR (CDCl_3) δ 171.4 (COCH_3), 169.2 (C-1), 113.1 (C), 100.5 (C-8), 84.8 (C-7), 83.9 (C-5), 79.4 (C-6), 66.2 (C-4), 62.1 (CH_2CH_3), 51.9 (C-2), 42.7 (C-3), 27.7 (CH_2), 27.6 (CH_2), 25.9 (CH_3), 24.8 (CH_3), 24.4 (CH_2), 21.0 (COCH_3), 13.9 (CH_2CH_3). Anal. calcd for $\text{C}_{11}\text{H}_{16}\text{O}_9\text{S}$; C, 49.52; H, 6.47. Found C, 49.28; H, 6.59. **Compound 9**: $[\alpha]_{\text{D}}^{25} -91.3$ (c 1.0, CHCl_3). ^1H NMR (CDCl_3) δ 6.17 (brs, 1H, H-1), 4.86 (dd, 1H, $J_{5,6} = 3.5$, $J_{6,7} = 6.0$ Hz, H-6), 4.89 (ddd, 1H, $J_{3a,4} = 2.0$, $J_{3b,4} = <0.5$, $J_{4,5} = 6.0$ Hz, H-4), 4.71 (brd, 1H, H-7), 4.24 (dd, 1H, H-5), 3.96–3.91 (m, 1H, SCH_2), and 3.48–3.39 (m, 1H, SCH_2), 2.66–2.57 (m, 2H, CH_2), 2.50–2.48 (m, 2H, CH_2 , H-3), 2.15–2.08 (m, 1H, CH_2), 2.08 (s, 3H, Ac), 1.95–1.87 (m, 1H, CH_2), 1.47 and 1.32 (s, 3H each, CH_3). ^{13}C NMR (CDCl_3) δ 172.6 (C-1), 169.1 (COCH_3), 113.5 (C), 100.5 (C-8), 84.9 (C-7), 81.9 (C-5), 79.0 (C-6), 73.2 (C-4), 44.3 (C-2), 40.4 (C-3), 27.0 and 26.9 ($-\text{SCH}_2-$ each), 25.8 and 24.4 (CH_3 , each), 24.1 ($-\text{CH}_2-$), 21.0 (COCH_3). Anal. calcd for $\text{C}_{11}\text{H}_{16}\text{O}_9\text{S}$; C, 49.21; H, 5.68. Found C, 49.16; H, 5.67.

3-Deoxy-6,7-*O*-isopropylidene-*D*-manno-2-octulosonic acid γ -lactone, cyclic 2-(1,3-propanediyl dithioacetal) (10)

To a solution of compound **9** (243 mg, 0.70 mmol) in methanol (10 mL) was added a solution of sodium methoxide in methanol (0.45 M, 0.1 mL) at rt. After stirring for 12 h, sodium borohydride (106 mg, 2.8 mmol) was added to the mixture at 0°C. After stirring for 2 h, the reaction mixture was neutralized with acetic acid and concentrated to a syrup. Purification by flash column chromatography (ethyl acetate:methanol = 9:1) of the residue afforded **10** as a colorless amorphous solid (210 mg, 86%).

$[\alpha]_{\text{D}}^{25} -13.0$ (c 1.0, CHCl_3). ^1H NMR (CDCl_3) δ 4.61 (ddd, 1H, $J_{3a,4} = 6.5$, $J_{3b,4} = 8.5$, $J_{4,5} = 8.0$ Hz, H-4), 4.39 (dd, 1H, $J_{5,6} = <0.5$, $J_{6,7} = 7.5$ Hz, H-6), 4.30 (ddd, 1H, H-7), 3.94 (dd, 1H, $J_{7,8a} = 4.0$, $J_{8a,8b} = 12.0$ Hz, H-8a), 3.82 (dd, 1H, $J_{7,8b} = <0.5$ Hz, H-8b), 3.90 and 2.60 (m, 1H each, $-\text{SCH}_2-$), 3.78 (dd, 1H, H-5), 3.46 and 2.64 (m, 1H each, $-\text{SCH}_2-$), 2.53 (ddd, 1H, $J_{3a,3b} = 14.0$ Hz, H-3b), 2.43 (ddd, 1H, H-3a), 2.19–2.25, and 1.85–1.96 (m, 1H each, $-\text{SCH}_2\text{CH}_2-$), 1.52 (s, 3H, CH_3), 1.39 (s, 3H, CH_3). ^{13}C NMR (CDCl_3) δ 173.1 (C-1), 108.6 (C), 76.6 (C-7), 76.0 (C-4), 75.5 (C-6), 70.5 (C-5), 60.6 (C-8), 44.5 (C-2), 40.8 (C-3), 27.0 (SCH_2-), 26.8 (SCH_2-), 26.4 (CH_3), 24.6 (SCH_2CH_2-), 24.1 (CH_3). Anal. calcd for $\text{C}_{14}\text{H}_{22}\text{O}_6\text{S}_2$; C, 47.98; H, 6.33. Found C, 47.82; H, 6.41.

Ammonium 3-deoxy-2-keto-*D*-manno-octulosonate (1a)

To a solution of the diol **10** (100 mg, 0.28 mmol) in 5.0 mL of the mixed solution (acetone/ $\text{H}_2\text{O} = 1/1$) was added *N*-bromosuccinimide (152 mg, 0.85 mmol)

at 0°C. The mixture was stirred for 30 min and then 1.0 mL of 5% Na₂S₂O₃ aq. was added. After removal of the acetone, the mixture was diluted with chloroform and separated. The aqueous phase was extracted with chloroform and the combined extracts were washed with brine, dried over Na₂SO₄, and concentrated. The residue was chromatographed on a column of silica gel (CHCl₃:MeOH = 2:1) to afford an amorphous syrup, and was treated with 90% aqueous trifluoroacetic acid (1 mL) at rt for 15 min. The mixture was concentrated under nitrogen, diluted with water, and neutralized by ammonia. The solution was passed through a Dowex 1 × 8-200 ion exchange resin. The combined filtrates were evaporated, and the residue was crystallized from ethanol-water to give 50 mg (70%) of colorless ammonium salt. m.p. 121–123°C; $[\alpha]_D^{25} + 39.2$ (c 1.0, H₂O).^[4g]

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REFERENCES

- For chemical syntheses of Kdo from D-arabinose, see: (a) Hershberger, C.; Binkley, S.B. Chemistry and metabolism of 3-deoxy-D-mannoctulosonic acid. I. Stereochemical determination. *J. Biol. Chem.* **1968**, *243*, 1578–1584. (b) Hershberger, C.; Davis, M.; Binkley, S.B. Chemistry and metabolism of 3-deoxy-D-mannoctulosonic acid. II. Practical synthesis and stability. *J. Biol. Chem.* **1968**, *243*, 1585–1588. (c) Cherniak, R.; Jones, R.G.; Gupta, D.S. Equilibria of the anomeric and ring forms of ammonium 3-deoxy-D-manno-octulosonate in deuterium oxide. *Carbohydr. Res.* **1979**, *75*, 39–49. (d) Schmidt, R.R.; Betz, R. Synthesis of 3-deoxy-D-manno-2-octulosonic acid (KDO). *Angew. Chem. Int. Ed. Engl.* **1984**, *23*, 430–431. (e) McNicholas, P.A.; Batley, M.; Redmond, J.W. Synthesis of methyl pyranosides and furanosides of 3-deoxy-D-manno-oct-2-ulosonic acid (KDO) by acid-catalysed solvolysis of the acetylated derivatives. *Carbohydr. Res.* **1986**, *146*, 219–231. (f) Dondoni, A.; Fantin, G.; Fogagnolo, M.; Merino, P.A. Concise new route to 3-deoxy-D-manno-2-octulosonic acid (KDO) from D-arabinose and 2-acetylthiazole. *Tetrahedron Lett.* **1990**, *31*, 4513–4516. (g) Smith, D.B.; Wang, Z.; Schreiber, S.L. The asymmetric epoxidation of divinyl carbinols: theory and applications. *Tetrahedron* **1990**, *46*, 4793–4808. (h) Abel, S.; Linker, T.; Giese, B. Ring opening of C-glycosides under mild conditions. *Synlett.* **1991**, 171–172. (i) Giese, B.; Carbone, B.; Göbel, T.; Muhn, R.; Wetterich, F. A new route to 3-deoxy-D-manno-2-octulosonic acid: Felkin-Anh rule in radical chemistry. *Tetrahedron Lett.* **1992**, *33*, 2673–2676. (j) Gao, J.; Härter, R.; Gordon, D.M.; Whitesides, G.M. Synthesis of KDO using indium-mediated allylation of 2,3:4,5-di-O-isopropylidene-D-arabinose in aqueous media. *J. Org. Chem.* **1994**, *59*, 3714–3715. (k) Sarabia, F.; Lopez-Herrera, F.J. β -Oxy- α -diazo-carbonyl compounds. Part 6: synthesis and reactivity of 3-deoxy-2-diazo-esters derived from monosaccharides. Applications in the synthesis of 2-C-alkyl-KDO analogues and related compounds. *Tetrahedron Lett.* **2001**, *42*, 8805–8809. (l) Li, L.S.; Wu, Y.L. Synthesis of 3-deoxy-2-ulosonic acid KDO and 4-epi-KDN, a highly efficient approach of 3-C homologation by propargylation and oxidation. *Tetrahedron* **2002**, *58*, 9049–9054. (m) Lichtenthaler, F.W.; Klotz, J.; Nakamura, K. (-)-Daucic acid: proof of D-*lyxo* configuration, synthesis of its D-*ribo*, D-*xylo*,

L-arabino and *L-lyxo* analogs, and biosynthetic implications. *Tetrahedron Asymm.* **2003**, *14*, 3973–3986. (n) Grison, C.; Petek, S.; Coutrot, P. Preparation of magnesium enolates of α -keto esters and synthetic applications. *Synlett.* **2005**, 331–333.

2. For chemical syntheses of Kdo from D-mannose, see: (a) Danishefsky, S.J.; Pearson, W.H.; Segmuller, B.E. Total synthesis of (+)-3-deoxy-D-manno-2-octulopyranosate. *J. Am. Chem. Soc.* **1985**, *107*, 1280–1285. (b) Hu, Y.J.; Huang, X.D.; Yao, Z.J.; Wu, Y.L. Formal synthesis of 3-deoxy-D-manno-2-octulosonic acid (KDO) via a highly double-stereoselective hetero Diels-Alder reaction directed by a (Salen)Co-II catalyst and chiral diene. *J. Org. Chem.* **1998**, *63*, 2456–2461. (c) Lubineau, A.; Augé, J.; Lubin, N. New strategy in the synthesis of 3-deoxy-D-manno-2-octulosonic acid (KDO), 2-deoxy-KDO and thioglycoside of KDO. *Tetrahedron* **1993**, *49*, 4639–4650. (d) Sugisaki, C.H.; Ruland, Y.; Le Clezio, I.; Baltas, M. New approaches in the synthesis of ulosonic acids and analogues. *Synlett.* **2001**, 1905–1908. (e) Sarabia, F.; Lopez-Herrera, F.J. β -Oxy- α -diazo-carbonyl compounds. Part 5: an improved synthesis of β -hydroxy- α -diazo esters derived from monosaccharides and synthetic applications in the chemistry of 3-deoxy-2-ulosonic acids. *Tetrahedron Lett.* **2001**, *42*, 8801–8804. (f) Barco, A.; Lucio, B.; Benetti, S.; Bertolasi, V.; De Risi, C.; Marchetti, P.; Pollini, G.P. 4-[(4-Methylphenyl)sulfonyl]-1-(triphenylphosphoranylidene)-2-butanone as a convenient precursor for a new formal synthesis of KDO. *Tetrahedron* **2002**, *58*, 8553–8558. (g) Hartmann, K.; Kim, B.G.; Linker, T. Manganese(III)-mediated radical reactions in carbohydrate chemistry: a new route to 3-deoxy-D-manno-oct-2-ulosonic acid (KDO). *Synlett.* **2004**, 2728–2731. (h) Kim, B.G.; Schilde, U.; Linker, T. New radical approaches to 3-deoxy-D-oct-2-ulosonic acids (KDO). *Synthesis* **2005**, 1507–1513. (i) Hekking, K.F.; Moelands, M.A.; van Delft, F.L.; Rutjes, F.P. An in-depth study on ring-closing metathesis of carbohydrate-derived α -alkoxyacrylates: efficient syntheses of DAH, KDO, and 2-deoxy-beta-KDO. *J. Org. Chem.* **2006**, *71*, 6444–6450.

3. For enzymatic syntheses, see: (a) Guerard, C.; Demuynek, C.; Bolte, J. Enzymatic synthesis of 3-deoxy-D-manno-2-octulosonic acid and analogues: a new approach by a non metabolic pathway. *Tetrahedron Lett.* **1999**, *40*, 4181–4182. (b) Sheftyan, G.Y.; Howe, D.L.; Wilson, T.L.; Woodard, R.W. Enzymatic synthesis of 3-deoxy-D-manno-octulosonate 8-phosphate, 3-deoxy-D-altro-octulosonate 8-phosphate, 3,5-dideoxy-D-gluco(manno)-octulosonate 8-phosphate by 3-deoxy-D-arabino-heptulosonate 7-phosphate synthase. *J. Am. Chem. Soc.* **1998**, *120*, 11027–11032.

4. (a) Charon, D.; Sarfati, R.S.; Strobach, D.R.; Szabó, L. Chemical synthesis and quantitative estimation of 3-deoxy D-manno-octulosonic acid. *Eur. J. Biochem.* **1969**, *11*, 364–369. (b) Paquet, F.; Sinaÿ, P. New stereocontrolled approach to 3-deoxy-D-manno-2-octulosonic acid containing disaccharides. *J. Am. Chem. Soc.* **1984**, *106*, 8313–8315. (c) Imoto, M.; Kusumoto, S.; Shiba, T. A new synthesis of 3-deoxy-D-manno-2-octulosonic acid (KDO) from D-mannose. *Tetrahedron Lett.* **1987**, *28*, 6235–6238. (d) Horito, S.; Amano, M.; Hashimoto, H. A new synthesis of 3-deoxy-D-manno-octulosonic acid (KDO). *J. Carbohydr. Chem.* **1989**, *8*, 681–684. (e) van der Klein, P.A.M.; Boons, G.J.P.H.; Veeneman, G.H.; van der Marel, G.A.; van Boom, J.H. An efficient route to 3-deoxy-D-manno-2-octulosonic acid (KDO) derivatives via a 1,4-cyclic sulfate approach. *Tetrahedron Lett.* **1989**, *30*, 5477–5480. (f) Ramage, R.; MacLeod, A.M.; Rose, G.W. Dioxalanones as synthetic intermediates. Part 6. Synthesis of 3-deoxy-D-manno-2-octulosonic acid (KDO), 3-deoxy-D-arabino-2-heptulosonic acid (DAH) and 2-keto-3-deoxy-D-gluconic acid (KDG). *Tetrahedron* **1991**, *47*, 5625–5636. (g) Coutrot, P.; Grison, C.; Tabyaoui, M. A new synthesis of 3-deoxy-D-manno-2-octulosonic acid. *Tetrahedron Lett.* **1993**, *34*, 5089–5092. (h) Tsukamoto, H.; Takahashi, T. New route to 3-deoxy-2-ulosonic acids; total syntheses of KDO and KDN. *Tetrahedron Lett.* **1997**, *38*, 6415–6418. (i) Reiner, M.; Schmidt, R.R. Synthesis of 3-deoxy-D-manno-2-octulosonic acid (Kdo) and D-glycero-D-talo-2-octulosonic acid (Ko) and their α -glycoside. *Tetrahedron Asymm.* **2000**, *11*, 319–335. (j) Mlynarski, J.; Banaszek, A. Synthetic approach to

- 3-deoxy-D-manno-oct-2-ulosonic acid (Kdo) α -disaccharides via a ketene dithioacetal. *Tetrahedron Asymm.* **2000**, *11*, 3737–3746. (k) Kumaran, G.; Mootoo, D.R. *Tetrahedron Lett.* **2001**, *42*, 3787–3785. (l) Hekking, K.F.W.; van Delft, F.L.; Rutjes, F. Ring-closing metathesis of α -ester-substituted enol ethers: application to the shortest synthesis of KDO. *Tetrahedron* **2003**, *59*, 6751–6758. (m) Kuboki, A.; Tajimi, T.; Tokuda, Y.; Kato, D.I.; Sugai, T.; Ohira, S. Concise synthesis of 3-deoxy-D-manno-oct-2-ulosonic acid (KDO) as a protected form based on a new transformation of α , β -unsaturated ester to α -oxocarboxylic acid ester via diol cyclic sulfite. *Tetrahedron Lett.* **2004**, *45*, 4545–4548. (n) Tanaka, H.; Takahashi, D.; Takahashi, T. Stereoselective synthesis of oligo- α (2,8)-3-deoxy-D-manno-2-octulosonic acid derivatives. *Angew. Chem. Int. Ed. Engl.* **2006**, *45*, 770–773.
5. Gurlain, T.; Wadouachi, A.; BeauPère, D. A new 6-C-alkylation from an alkyl mannofuranoside 5,6-cyclic sulfate. *Carbohydr. Res.* **2000**, *324*, 66–73.
6. a) Brady, T.P.; Kim, S.H.; Wen, K.; Kim, C.; Theodorakis, E.A. Norrisolide: total synthesis and related studies. *Chem. Eur. J.* **2005**, *11*, 7175–7190. (b) Schmidt, O.T. Isopropylidene Derivatives. *Methods Carbohydr. Chem.* **1963**, *2*, 319.
7. Horton, D.; Jewell, J.S. 1,6-anhydro-2,3-*O*-isopropylidene- β -D-lyxo-hexopyranos-4-ulose: studies on oxidation of carbohydrate derivatives. *Carbohydr. Res.* **1966**, *2*, 251–260.
8. Garcia-Moreno, M.I.; Mellet, C.O.; Fernandez, J.M.G. Synthesis of calystegine B2, B3, and B4 analogues: mapping the structure-glycosidase inhibitory activity relationships in the 1-deoxy-6-oxacalystegine series. *Eur. J. Org. Chem.* **2004**, 1803–1819.
9. Gao, Y.; Sharpless, K.B. Vicinal diol cyclic sulfates: like epoxides only more reactive. *J. Am. Chem. Soc.* **1988**, *110*, 7538–7539.
10. Gelas, J.; Horton, D. Kinetic acetonation of D-mannose: preparation of 4,6-mono- and 2,3:4,6-di-*O*-isopropylidene-D-mannopyranose. *Carbohydr. Res.* **1978**, *67*, 371–387.
11. Yamasaki, R.; Koshino, H.; Kurono, S.; Nishinaka, Y.; McQuillen, P.A.; Kumi, A.; Gulati, S.; Rice, P.A. Structural and immunochemical characterization of a *Neisseria gonorrhoeae* epitope defined by a monoclonal antibody 2C7; the antibody recognizes a conserved epitope on specific lipo-oligosaccharides in spite of the presence of human carbohydrate epitopes. *J. Biol. Chem.* **1999**, *274*, 36550–36558.
12. Kubo, H.; Ishii, K.; Koshino, H.; Toubetto, K.; Naruchi, K.; Yamasaki, R. Synthesis of a 3,4-di-*O*-substituted heptose structure: a partial oligosaccharide expressed in neisserial lipooligosaccharide. *Eur. J. Org. Chem.* **2004**, 1202–1213.
13. Ishii, K.; Esumi, Y.; Iwasaki, Y.; Yamasaki, R. Synthesis of a 2,3-di-*O*-substituted heptose structure by regioselective 3-*O*-silylation of a 2-*O*-substituted heptose derivative. *Eur. J. Org. Chem.* **2004**, 1214–1227.